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SCREENING PROCEDURE FOR DETECTION OF ANTIDEPRESSANTS AND THEIR METABOLITES IN URINE USING A COMPUTERIZED GAS CHROMATOGRAPHIC—MASS SPECTROMETRIC TECHNIQUE*

HANS MAURER and KARL PFLEGER*

*Institut für Pharmakologie und Toxikologie der Universität des Saarlandes,
D-6650 Homburg/Saar (F.R.G.)*

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SUMMARY

A method for the identification of antidepressants and their metabolites in urine after acid hydrolysis is described. The acetylated extract is analysed by computerized gas chromatography—mass spectrometry. An on-line computer allows rapid detection using mass fragmentography with the masses 58, 84, 86, 100, 191, 193, 194, 205. The identity of positive signals in the reconstructed mass fragmentogram is established by a comparison of the entire mass spectra with those of standards. The mass fragmentogram, the underlying mass spectra and the gas chromatographic retention indices (OV-101) are documented.

INTRODUCTION

Within the scope of a screening procedure for detection of psychotropic and addictive drugs and their metabolites in urine [1, 2], screening for benzodiazepines [3], butyrophenone and bisfluorophenyl neuroleptics [4, 5], anti-inflammatory analgesics [6] and opioids and other potent analgesics [7] has been described; screening for phenothiazine and analogous neuroleptics and antiparkinsonian drugs is in preparation [8]. Screening for tri- and tetracyclic antidepressants is described below. Such a screening is necessary in analytical toxicology to diagnose a probable intoxication. Furthermore, antidepressants are encountered frequently in analysis, when monitoring patients who may have taken addictive drugs and simultaneously taken antidepressants therapeutically. Only a gas chromatographic—mass spectrometric technique (GC—MS) detecting a few of these drugs as parent compounds in gastric

*These results were reported in part at the Jahrestagung der Deutschen Gesellschaft für klinische Chemie, Stuttgart, F.R.G., September 23th and 24th, 1982.

contents or plasma after intoxications has been documented [9]. The following procedure allows rapid identification and differentiation of 21 antidepressants and their metabolites in urine after therapeutic dosage. If necessary, plasma levels of the identified drugs can be determined using a procedure (e.g. gas-liquid chromatography, high-performance liquid chromatography, radio-immunoassay) described and cited in the review article of Scoggins et al. [10].

EXPERIMENTAL

Apparatus, sample preparation (acid hydrolysis, extraction, acetylation) and the GC-MS technique used for this study have been previously described [3, 5, 6, 11].

RESULTS AND DISCUSSION

All investigations were carried out using the urine of man with the exception of dimetacrine, melitracene, noxyptiline, protriptyline and trazodone, which were detected (in the absence of human samples) in the urine of rats. Some of the antidepressants are excreted in urine completely metabolized and conjugated. Therefore, the conjugates were decomposed by acid hydrolysis, which can be completed more quickly than enzymatic hydrolysis. To improve their GC characteristics hydroxy and amino groups were acetylated.

The results of our investigations are shown in Table I. The mass fragmentogram with the eight proposed masses allows the detection of 21 antidepressants and their metabolites. Some of these compounds are acetylated. The retention indices were determined using a gas chromatograph combined with a flame ionisation detector and a nitrogen-sensitive flame ionisation detector with a temperature programme [3]. In our experience retention indices give preliminary indications and may be useful to gas chromatographers without a GC-MS facility and so they are given here. Furthermore, they allow one to distinguish between the isomeric hydroxy metabolites which give the same mass spectra (mass spectra Nos. 9, 24 and 26 in Fig. 1). All of the mass spectra are shown in Fig. 1 for the precise identification of the compounds. Formulae are proposed for probable structures of metabolites.

Only those metabolites that were usually found are listed. Because of individual differences in metabolism, time elapsed after administration and different routes of administration, not all metabolites given in Table I were detected in each sample. Further metabolites can be found. The mass spectra of those will be included in a forthcoming handbook [12].

Dimetacrine and its metabolites are completely decomposed by the acid hydrolysis. Thus, they must be identified in a direct extract [11] of urine.

Amitriptyline, amitriptyline-N-oxide and nortriptyline lead to common metabolites. If amitriptyline-N-oxide was taken, amitriptyline and noramitriptyline (mass spectra Nos. 1 and 4) cannot be detected. If nortriptyline was taken, amitriptyline (mass spectra Nos. 1, 2 and 3) cannot be detected.

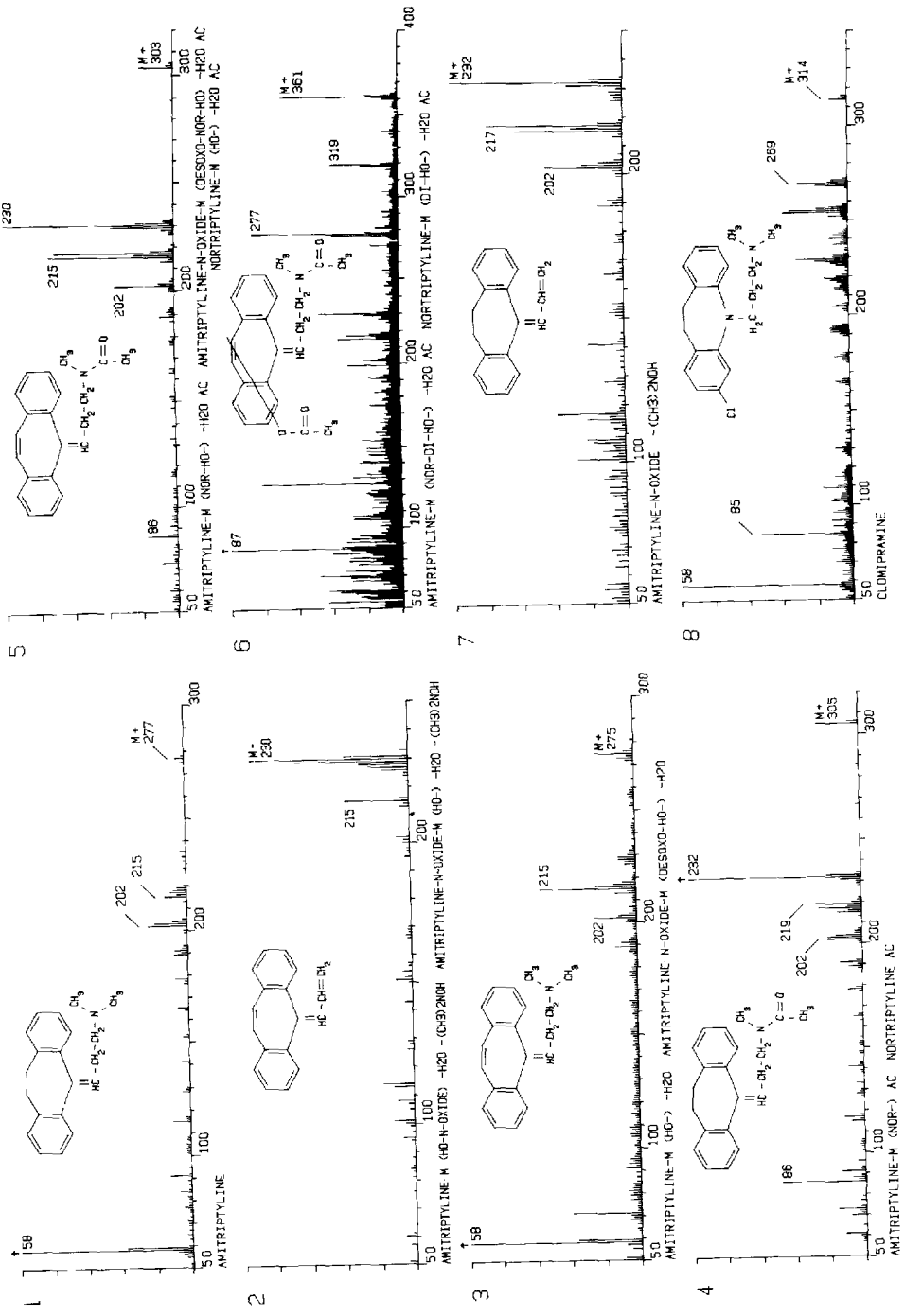
Desipramine, imipramine and lofepramine also lead to common metabolites. If desipramine or lofepramine were taken, imipramine and hydroxyimipramine (mass spectra Nos. 27 and 29) cannot be detected. Desipramine and lofe-

TABLE I

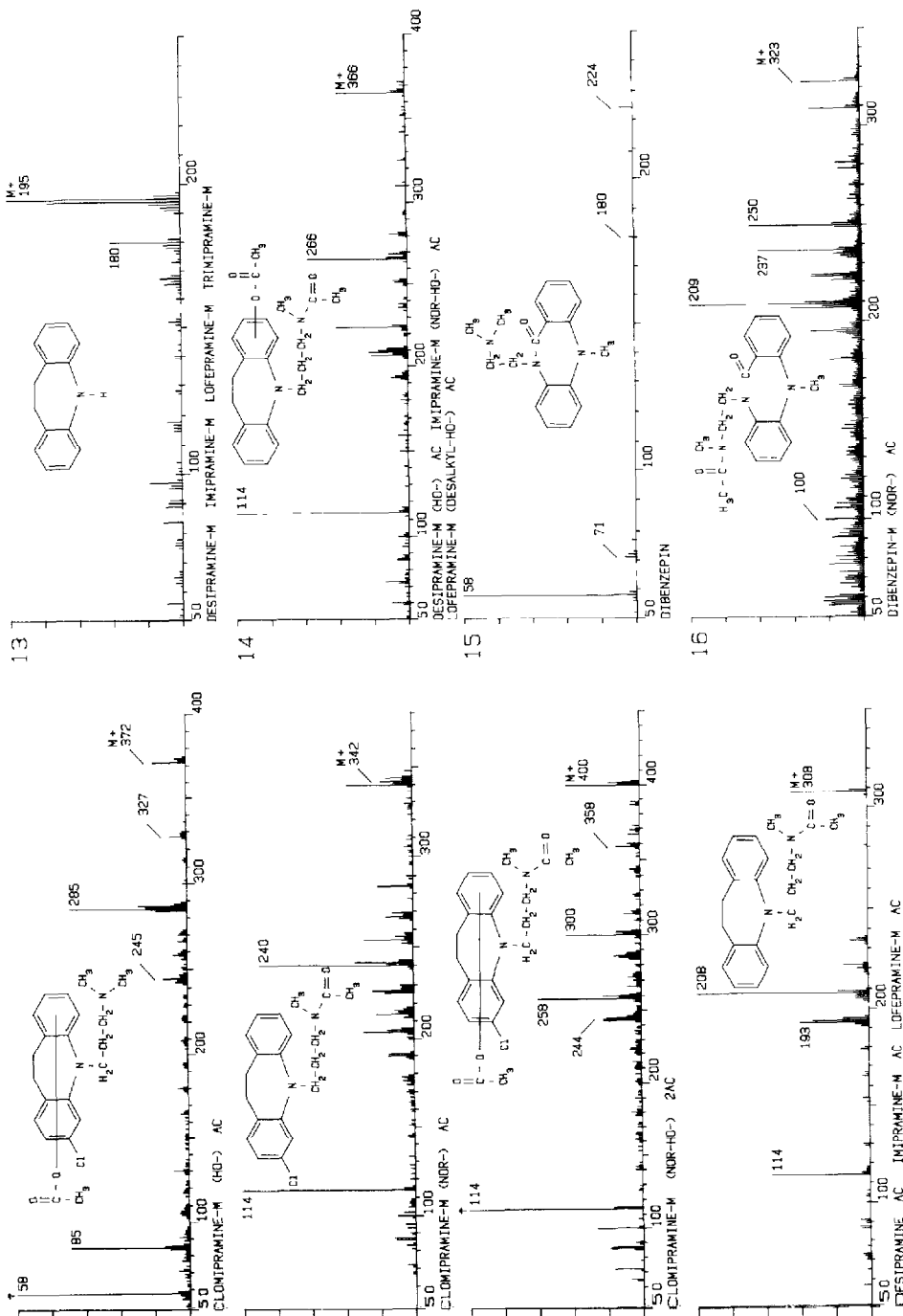
MONITORING PROGRAMME FOR ANTIDEPRESSANTS AND THEIR METABOLITES

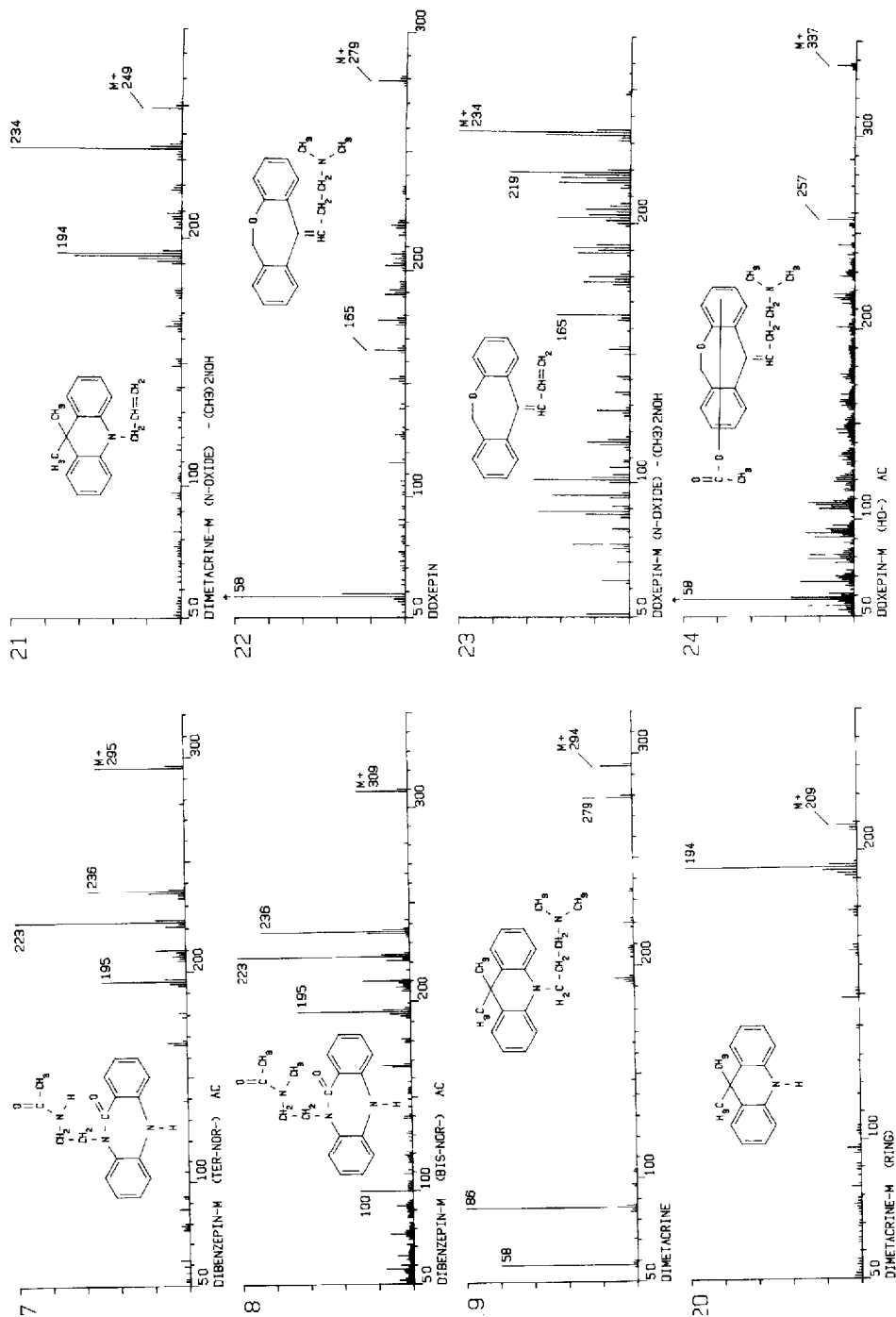
Mass spectrum No.	M ⁺ Drug/metabolite (M)	m/e (relative intensities)										Retention index (OV-101)
		58	84	86	100	191	193	194	205			
11	277 Amitriptyline	+++										2204
12	230 M (HO-N-oxide)											2000 FID
13	275 M (HO-)	+++										2236
14	305 M (Nor-)			+								2660
15	303 M (Nor-HO-)			+								2670
16	361 M (Nor-di-HO-)			+								2800
17	232 Amitriptyline-N-oxide											1976 FID
18	230 M (HO-)											2000 FID
19	275 M (Desoxo-HO-)	+++										2236
20	303 M (Desoxo-nor-HO-)			+								2670
21	314 Clomipramine	+++										2457
22	372 M (HO-) 1st isomer	+++										2806
23	372 M (HO-) 2nd isomer	+++										2906
24	342 M (Nor-)			+							+	2994
25	400 M (Nor-HO-)			+								3205
26	308 Desipramine	+		+					+	++	+	2669
27	195 M (ring)								+	+	++	1931
28	366 M (HO-)			+						+		3066
29	295* Dibenzepin	+++										2465
30	323 M (Nor-)								+	+	+	2800
31	295 M (Ter-nor-)								+	+	+	2824
32	309 M (Bis-nor-)	+			++					+	+	2869
33	294 Dimetacrine	++		+++								2313
34	209 M (ring)								+	+	+++	1906
35	249 M (N-oxide)								+	++	++	2020

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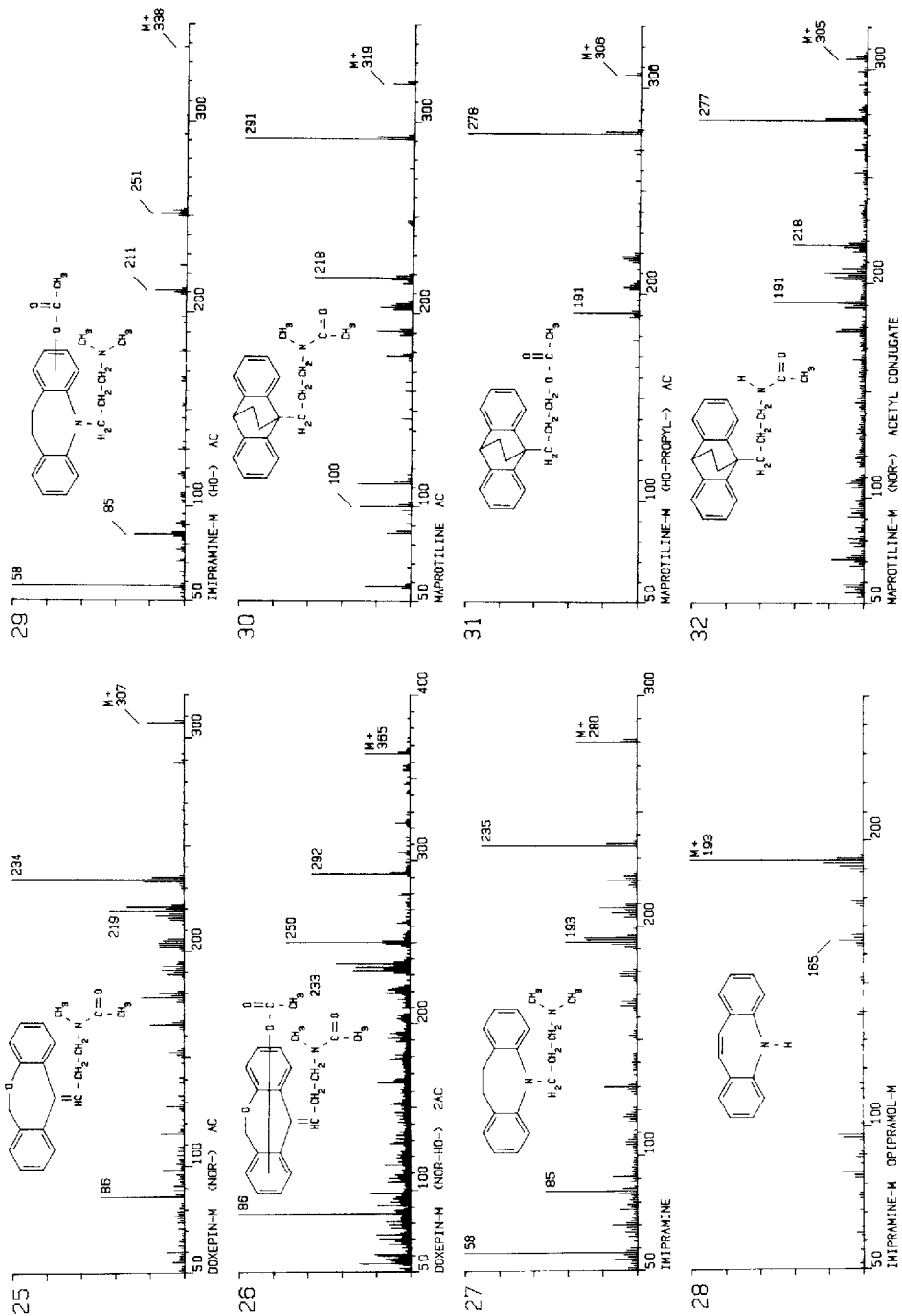
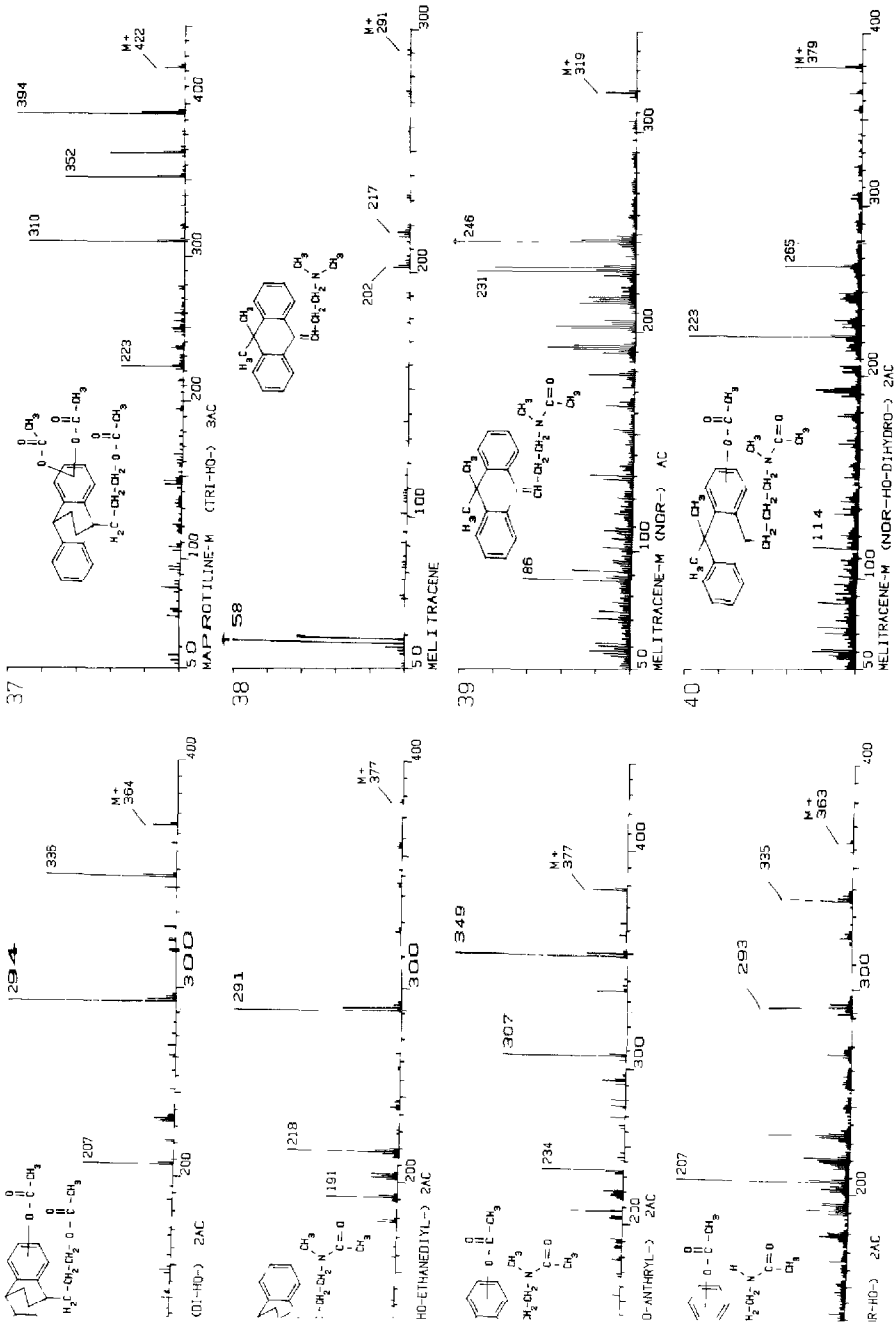


Fig. 1.



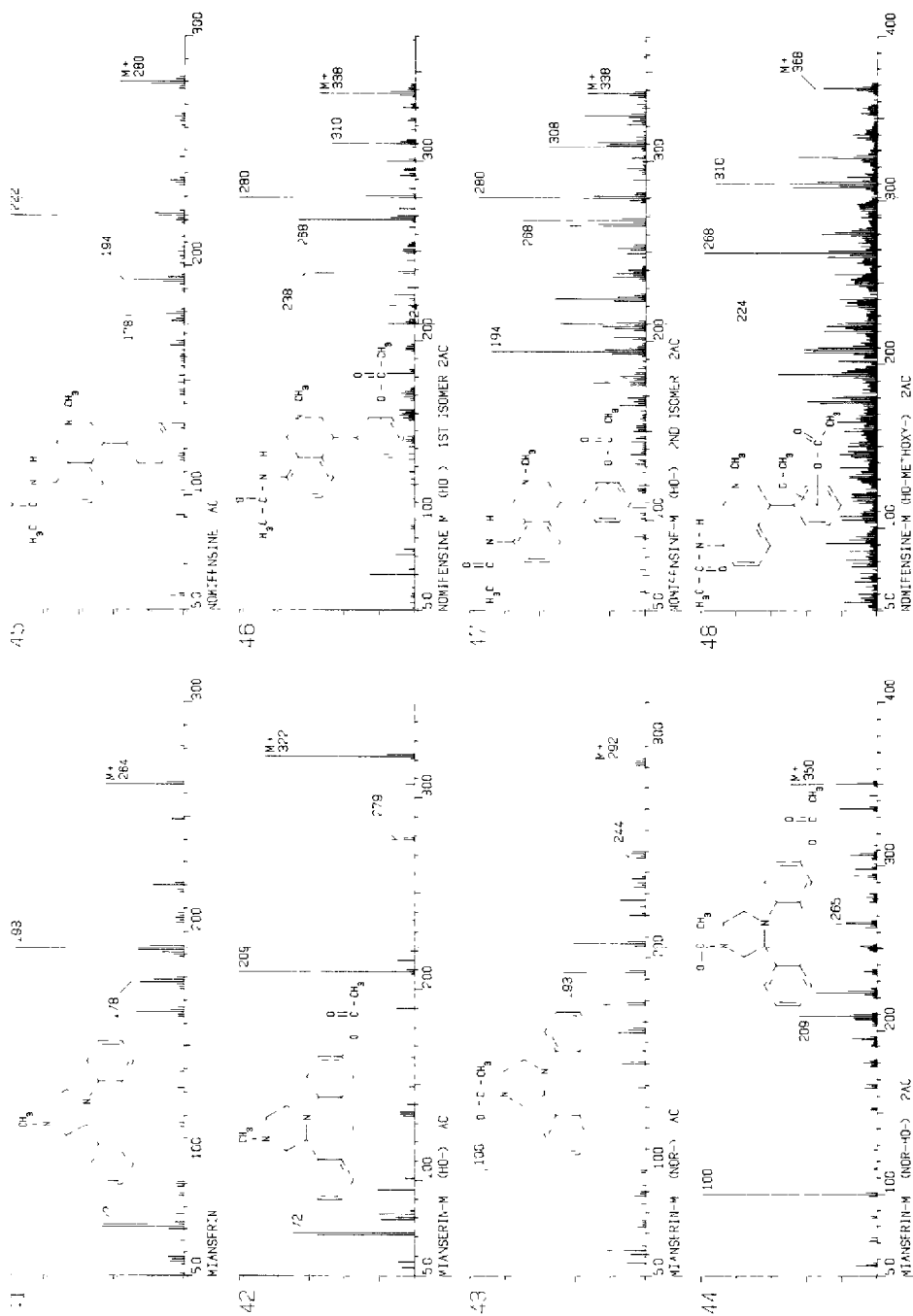
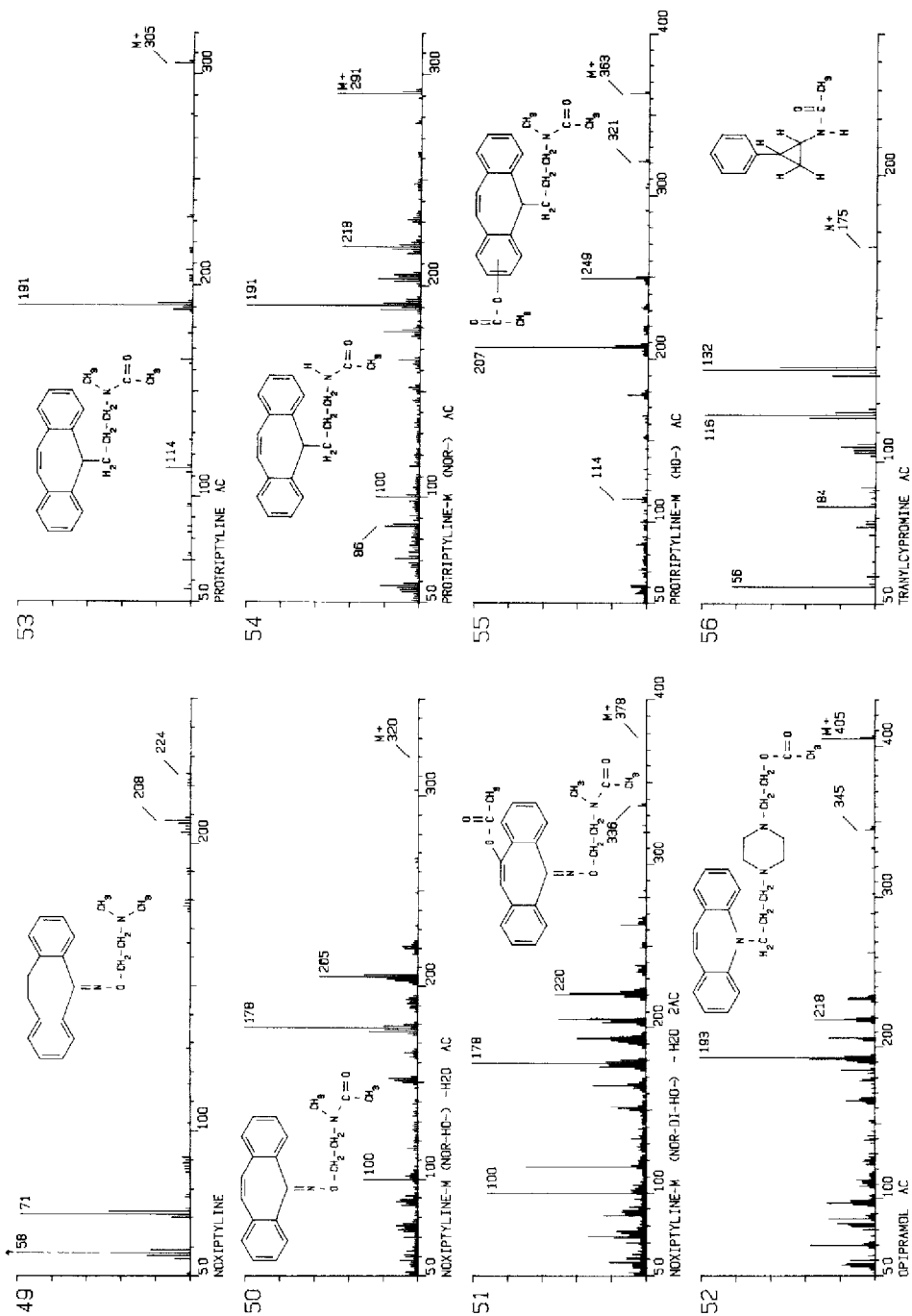


Fig. 1.

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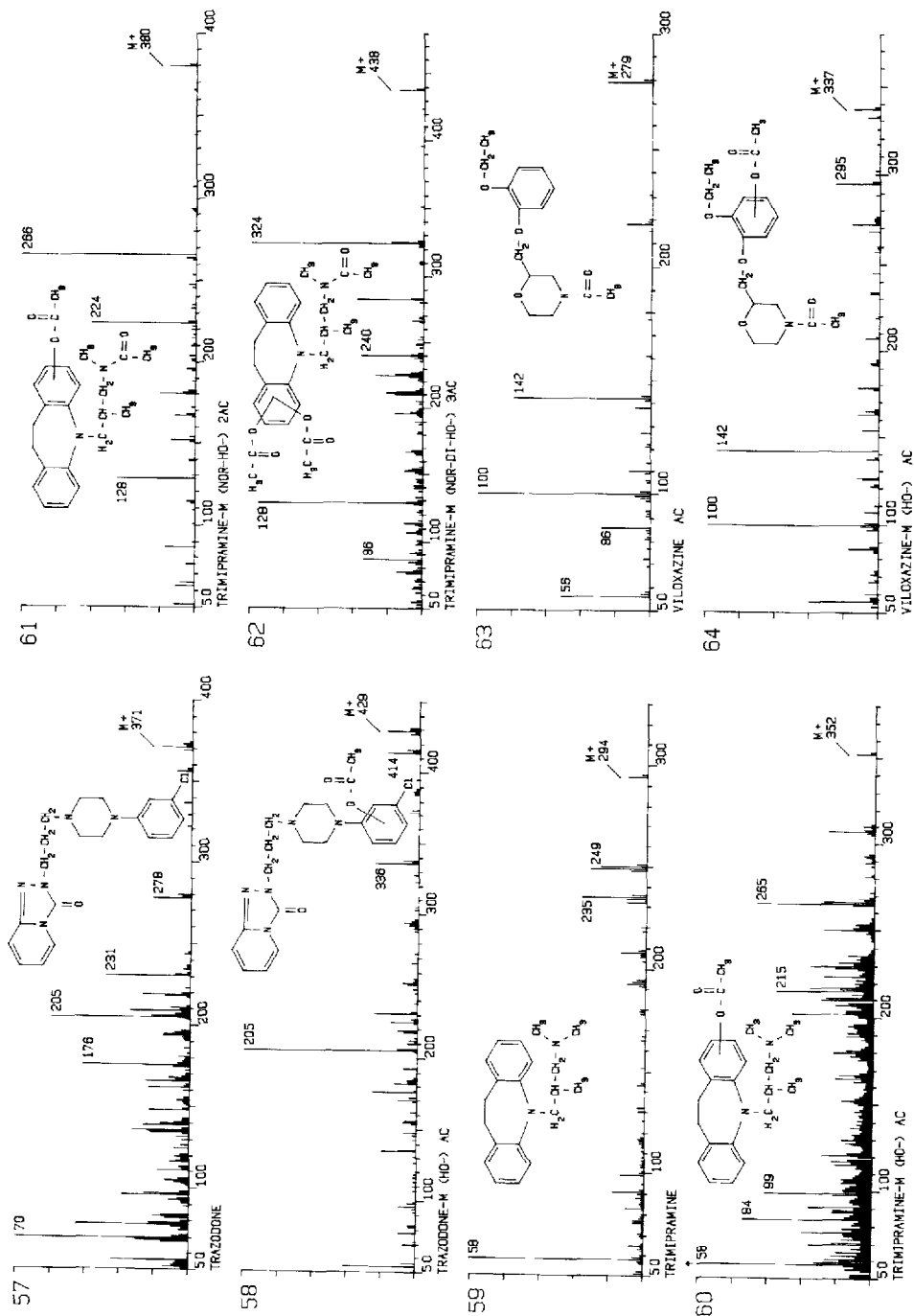


Fig. 1. Mass spectra of antidepressants and their metabolites identified in urine after hydrolysis, extraction and acetylation.

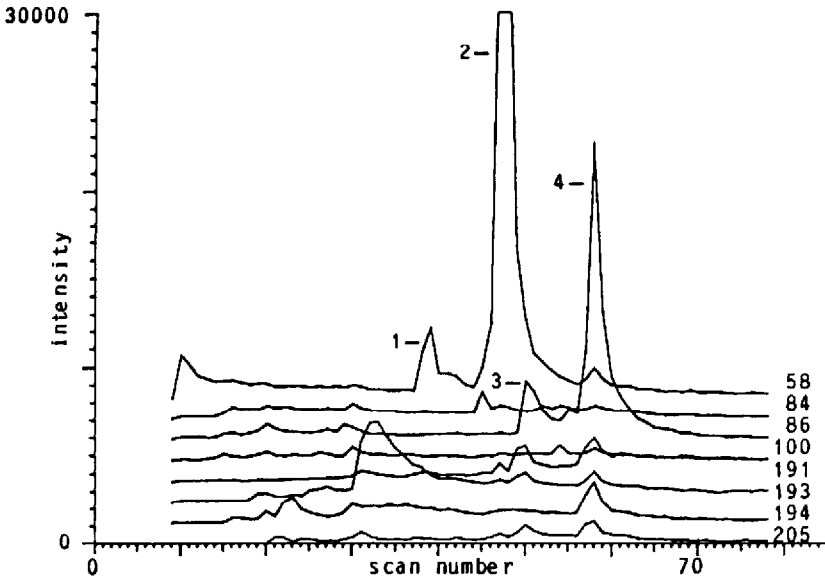


Fig. 2. Mass fragmentogram of doxepin and its metabolites. For identification of peaks, see text.

pramine cannot be differentiated by this screening procedure because lofepramine is not volatile under the GC conditions used. If necessary, the ingestion of lofepramine can be detected by identifying *p*-chlorobenzoic acid after a sample preparation described in the literature [13].

Two kinds of artifacts result from the analytical procedure used. Metabolites with alcoholic hydroxy groups eliminate water, and N-oxides undergo the Cope elimination reaction [14]. The mass spectra of these artifacts are included in Fig. 1 (mass spectra Nos. 2, 3, 5, 6, 50 and 51, and Nos. 2, 7, 21 and 23, respectively).

Because all compounds possibly indicated by the mass fragmentogram can be precisely differentiated by comparison of the underlying mass spectra with those of standards (Fig. 1), interference by other drugs is impossible. Furthermore, if the resolution of the peaks is imperfect, a temperature programme with a lower rate or an isothermic procedure can be used. Where there is still doubt a capillary column can help to obtain sufficient resolution.

To illustrate the method, a mass fragmentogram from a psychiatric patient is shown in Fig. 2. Peak 1 indicates doxepin (mass spectrum No. 22), peak 2 the two isomers of hydroxydoxepin (mass spectrum No. 24), peak 3 nordoxepin (mass spectrum No. 25) and peak 4 the two isomers of norhydroxydoxepin (mass spectrum No. 26). This example shows that the presented screening procedure allows a rapid and exact identification and differentiation of antidepressants and their metabolites in urine.

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